

The Synthesis of GABA_A Active Ligands by the Stille Process

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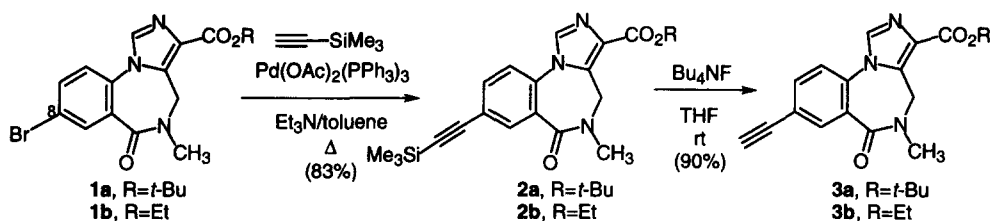
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Abstract: A variety of GABA_A active analogs **3b-3h** with different substituents at C(8) of the imidazobenzodiazepine nucleus was synthesized via the Stille coupling reaction. In addition, stannane **4** was prepared which provides facile entry into other members of the series. © 1997 Elsevier Science Ltd.

The GABA_A/benzodiazepine receptor (GABA_A/BzR) is a heterooligomeric family of ligand gated ion channels which constitutes the major inhibitory neurotransmitter system in the mammalian central nervous system.¹ This membrane-bound protein complex plays a central role in the molecular mechanisms which underlie anxiety, sleep, convulsions and cognition; consequently, it represents an important target for the design of selective agents to treat specific disease states in the CNS. Presently, a total of 6 α , 3 β , 3 γ , 1 δ and 2 ρ subunits, which have been isolated from various GABA_A receptor isoforms from the mammalian CNS, have been cloned and sequenced.¹ Among these, α 1 β 2 γ 2 represents the classical Type-I BzR, while α 2 β 2 γ 2, α 3 β 2 γ 2 and α 5 β 2 γ 2 ion channels are termed Type-II BzR.^{2,3} The closely related α 4 β 2 γ 2 and α 6 β 2 γ 2 channels both resemble "diazepam-insensitive" sites,⁴⁻⁶ the latter of which has been studied extensively.⁷⁻¹² The extensive molecular diversity of GABA_A/BzRs which results from these subunits has been implicated in the multiple pharmacological properties elicited by ligands which lack subtype selectivity, such as diazepam.^{1,13-16} Furthermore, the regional heterogeneity of GABA_A/BzR subtypes has also been suggested as a basis for the multiplicity of pharmacological properties of the benzodiazepines.^{1,16}

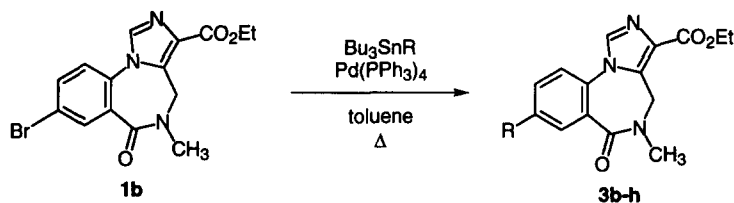
Recently, the synthesis of a series of C(8)-substituted imidazobenzodiazepines which exhibit high affinity ($K_i \approx 0.4-5$ nM) and selectivity for recombinant α 5-containing GABA_A receptors by a Heck-type coupling reaction¹⁷ was reported.^{18,19} For example, as illustrated in Scheme 1, the 8-bromobenzodiazepine templates **1a-b**⁷ were coupled with acetylenetriethylsilane in the presence of bis(triphenylphosphine)-palladium(II) acetate (10 mol %) to provide trimethylsilyl analogs **2a-b** in 83% yield. Treatment of **2a-b** with Bu₄NF effected the desilylation to furnish the 8-acetylenebenzodiazepines **3a-b** in 90% yield. Substitution of an acetylene unit at C(8) of the imidazobenzodiazepine nucleus has provided the most selective ligands reported to date for the α 5-containing GABA_A receptors (67-fold for **2a** and 60-fold for **2b**). The α 5-containing isoforms have been found principally in the hippocampus^{20,21} which is involved in memory and learning (cognition).²²⁻²⁴ Moreover, the action *in vivo* of these ligands resembled that of inverse agonists (convulsants/proconvulsants).

Scheme 1



In order to readily expand the SAR of these imidazobenzodiazepines at C(8), it was decided to prepare a number of analogs by utilization of the Stille coupling reaction.^{25,26} This process has been employed widely to functionalize an aromatic ring with vinyl, allyl or aryl groups *via* organotin compounds and Pd catalysts.^{25,26}

Scheme 2

Table 1. Stille coupling reactions of **1b** with tributyltin reagents.

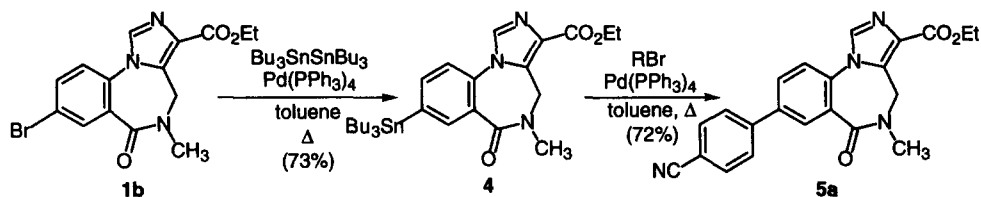
	R	time (h)	yield (%)
3b		6	50
3c		2	90
3d		4	95
3e		4	72
3f		4	83
3g		2	92
3h		3.5	94

As illustrated in Scheme 2, 8-bromobenzodiazepine **1b** was treated with a variety of tributyltin reagents in the presence of tetrakis(triphenylphosphine) palladium(0) as a catalyst. A typical reaction

procedure is illustrated here. A mixture of bromide **1b** (0.41 mmol, 1 eq.), tributyltin reagent (1.1 eq) and Pd(PPh₃)₄ (0.03-0.05 eq.) in dry toluene (5 mL) was heated to reflux under N₂ for a period of several hours (as shown in Table 1). The color of the solution changed from yellow to black and Pd(0) precipitated from the medium. The mixture was diluted with EtOAc (50 mL) and washed with a 15% aqueous solution of KF. The organic layer was separated and dried (K₂CO₃). After removal of the solvents, the residue was purified by flash chromatography (silica gel, EtOAc) to provide the C(8)-substituted imidazobenzodiazepines represented by **3**. A variety of vinyl, allyl and aryl tributyltin reagents underwent the Stille coupling reaction to give C(8)-substituted benzodiazepines **3b-h** in good to excellent yield (Table 1). The $\alpha 5$ selective ligand **3b** was also prepared in one step. Initial *in vitro* pharmacology indicated that vinyl analog **3c** exhibited very good affinity (K_i \approx 0.4 nM) and some selectivity (21-fold) for recombinant $\alpha 5$ -containing GABA_A receptors as compared to the acetylene analog **3b**.

In addition, as illustrated in Scheme 3, 8-stannylimidazobenzodiazepine **4** was prepared in 73% yield and characterized by heating bromide **1b** with dibutyltin in the presence of Pd(PPh₃)₄.²⁷ Stannane **4** is now available to react with a variety of common coupling partners including vinyl, allyl, aryl, and acyl halides as well as aryl and vinyl triflates to provide more analogs for the study of $\alpha 5$ containing GABA_A receptors. For example, stannane **4** was stirred with 4-bromobenzonitrile to provide **5a**.

Scheme 3



In summary, the Stille coupling reaction has been employed for the synthesis of a number of analogs with a variety substituents at C(8) of the imidazobenzodiazepine nucleus of **1b**. These reactions, as expected, occurred in good to excellent yield. In addition, the preparation of stannane **4** will provide facile entry into a host of important analogs for biological screening at GABA_A/Bz receptors.

Acknowledgment: The authors wish to thank the NIMH (MH46851) for generous financial support as well as Dr. Ruth M. McKernan (Merck, Sharp & Dohme Research Laboratories) and Dr. Phil Skolnick (NIH) for biological screening of these compounds.

References:

- (1) Sieghart, W. *Pharm. Rev.* **1995**, *47*, 181.
- (2) Benavides, J.; Peny, B.; Ruano, D.; Vitorica, J.; Scatton, B. *Brain Res.* **1993**, *604*, 240.
- (3) Mertens, S.; Benke, D.; Mohler, H. *J. Biol. Chem.* **1993**, *268*, 5965.
- (4) Sieghart, W.; Eichinger, A.; Richards, J. G.; Mohler, H. *J. Neurochem.* **1987**, *48*, 46.

- (5) Wisden, W.; Herb, A.; Wieland, H.; Keinanen, K.; Luddens, H.; Seeburg, P. H. *FEBS Lett.* **1991**, *289*, 227.
- (6) Gunnensen, G.; Kaufman, C.; Skolnick, P. *Neuropharmacology* in press,
- (7) Gu, Z. Q.; Wong, G.; Dominguez, C.; de-Costa, B. R.; Rice, K. C.; Skolnick, P. *J. Med. Chem.* **1993**, *36*, 1001.
- (8) Fryer, R. I.; Zhang, P.; Lin, K.-Y.; Upasani, R. B.; Wong, G.; Skolnick, P. *Med. Chem. Res.* **1993**, *3*, 183.
- (9) Wong, G.; Koehler, K. F.; Skolnick, P.; Gu, Z. Q.; Ananthan, S.; Schönholzer, P.; Hunkeler, W.; Zhang, W.; Cook, J. M. *J. Med. Chem.* **1993**, *36*, 1820.
- (10) Wong, G.; Gu, A.-Q.; Fryer, R. I.; Skolnick, P. *Med. Chem. Res.* **1992**, *2*, 217.
- (11) Wong, G.; Skolnick, P. *Eur. J. Pharmacol. Mol. Pharm. Sec.* **1992**, *225*, 63.
- (12) Zhang, P.; Zhang, W.; Liu, R.; Harris, B.; Skolnick, P.; Cook, J. M. *J. Med. Chem.* **1995**, *38*, 1679.
- (13) Whiting, J. G.; McKernan, R. M.; Iversen, L. L. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 9966.
- (14) Mohler, H.; Benke, D.; Mertens, S.; Fritschy, J. M. GABA_A-Receptor subtypes differing in α -subunit composition display unique pharmacological properties In *GABAergic Synaptic Transmission*; G. Biggio; A. Concas and E. Costa, Ed.; Raven Press: New York, 1992; p 41.
- (15) Hadingham, K. L.; Wingrove, P.; Le-Bourdelle, B.; Palmer, K. J.; Ragan, C. I.; Whiting, P. J. *Mol. Pharmacol.* **1993**, *43*, 970.
- (16) Doble, A.; Martin, I. L. *Trends Pharmacol. Sci.* **1992**, *13*, 76.
- (17) Austin, W. B.; Bilow, N.; Kelleghan, W. J.; Lau, K. S. Y. *J. Org. Chem.* **1981**, *46*, 2280.
- (18) Liu, R.; Zhang, P.; McKernan, R. M.; Wafford, K.; M., C. J. *Med. Chem. Res.* **1995**, *5*, 700.
- (19) Liu, R.; Hu, R. J.; Zhang, P.; Skolnick, P.; Cook, J. M. *J. Med. Chem.* **1996**, *39*, 1928.
- (20) Wisden, W.; Laurie, D. J.; Monyer, H.; Seeburg, P. H. *J. Neurosci.* **1992**, *12*, 1040.
- (21) McKernan, R. M.; Quirk, K.; Prince, R.; Cox, P. A.; Gillard, N. P.; Ragan, C. I.; Whiting, P. J. *Neuron* **1991**, *7*, 667.
- (22) Izquierdo, I.; Medina, J. H. *Trends Pharmacol. Sci.* **1991**, *12*, 260.
- (23) Izquierdo, I.; Medina, J. H.; Da-Cunha, C.; Wolfman, C.; Jerusalinsky, D.; Ferreira, M. B. *Braz. J. Med. Biol. Res.* **1991**, *24*, 865.
- (24) Werck, M. C.; Daval, J. L. *Pediatr. Res.* **1991**, *30*, 100.
- (25) Stille, J. K. *Angew. Chem. Int. Ed. Engl.* **1986**, *25*, 508.
- (26) Mitchell, T. N. *Synthesis* **1992**, 803.
- (27) **4**: ¹H NMR (250 MHz, CDCl₃) δ 0.86 (t, 9H, J = 7.2 Hz), 1.02-1.58 (m, 21H), 3.22 (s, 3H), 4.41 (q, 2H, J = 6.8 Hz), 4.43 (s, br, 1H), 5.14 (s, br, 1H), 7.32 (d, 1H, J = 7.7 Hz), 7.67 (d, 1H, J = 7.7 Hz), 7.85 (s, 1H), 8.10 (s, 1H); MS (CI, CH₄) *m/e* (relative intensity) 576 (M+1, 100).

(Received in USA 25 August 1997; revised 22 September 1997; accepted 24 September 1997)